
MuG - CHi-C Pipeline Documentation

Release 0.1

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Requirements and Installation

1.1 Requirements

1.1.1 Software

- Python 2.7.12+
- R >=3.1.2
- bedtools
- perl
- HiCUP
- bwa
- GEM
- TADbit
- samtools>1.3

1.1.2 Python Modules

- mg-tool-api
- pylint
- pytest
- pandas
- rtree

1.1.3 R Modules

- argparse
- devtools
- Chicago

To Run `runChicago.py` and `process_runChicago.py`, the R script `runChicago.R` from <https://bitbucket.org/chicagoTeam/chicago/src/ceffddda8ea392a1e84e4db9593f8fc35ac88048/chicagoTools/?at=master> should be downloaded and added to PATH.

1.2 Installation

Directly from GitHub:

```
git clone https://github.com/pabloacera/C-HiC.git
```

Using pip:

```
pip install git+https://github.com/pabloacera/C-HiC.git
```

Install R modules, use the following R code:

```
install.packages("argparser")      install.packages("devtools")      library(devtools)      in-  
stall_bitbucket("chicagoTeam/Chicago", subdir="Chicago")
```

CHAPTER 2

Full Installation

The following document is for the full installation of all software required by the C-HiC module and all programmes that it uses. The document has been written with Ubuntu Linux, although many of the commands are cross platform (*nix) compliant.

If you already have certain packages installed feel free to skip over certain steps. Likewise the bin, lib and code directories are relative to the home dir; if this is not the case for your system then make the required changes when running these commands.

2.1 Setup the System Environment

```
1 sudo apt-get install -y make build-essential libssl-dev zlib1g-dev      \\  
2 libbz2-dev libreadline-dev libsqlite3-dev wget curl llvm libncurses5-dev \\  
3 libncursesw5-dev xz-utils tk-dev unzip mcl libgtk2.0-dev r-base-core    \\  
4 libcurl4-gnutls-dev python-rpy2 git libtbb2 pigz liblzma-dev libhdf5-dev \\  
5 texlive-latex-base  
6  
7 cd ${HOME}  
8 mkdir bin lib code  
9 echo 'export PATH="${HOME}/bin:$PATH"' >> ~/.bash_profile
```

2.2 Setup pyenv and pyenv-virtualenv

This is required for managing the version of Python and the installation environment for the Python modules so that they can be installed in the user space.

```
1 git clone https://github.com/pyenv/pyenv.git ~/.pyenv  
2 echo 'export PYENV_ROOT="${HOME}/.pyenv"' >> ~/.bash_profile  
3 echo 'export PATH="${PYENV_ROOT}/bin:$PATH"' >> ~/.bash_profile  
4 echo 'eval "$(pyenv init -)"' >> ~/.bash_profile
```

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```
5
6 # Add the .bash_profile to your .bashrc file
7 echo 'source ~/.bash_profile' >> ~/.bashrc
8
9 git clone https://github.com/pyenv/pyenv-virtualenv.git ${PYENV_ROOT}/plugins/pyenv-
  ↳ virtualenv
10
11 pyenv install 2.7.12
12 pyenv virtualenv 2.7.12 C-HiC
13
14 # Python 3 environment required by iNPS
15 pyenv install 3.5.3
16 ln -s ${HOME}/.pyenv/versions/3.5.3/bin/python ${HOME}/bin/py3
```

2.3 Installation Process

2.3.1 bedtools and libspatialindex-dev

```
1 sudo apt-get install bedtools
2 sudo apt-get install libspatialindex-dev
```

2.3.2 Bowtie2 Aligner

```
1 cd ${HOME}/lib
2 wget --max-redirect 1 https://downloads.sourceforge.net/project/bowtie-bio/bowtie2/2.
  ↳ 3.4/bowtie2-2.3.4-linux-x86_64.zip
3 unzip bowtie2-2.3.4-linux-x86_64.zip
```

2.3.3 HiCUP

```
1 cd ${HOME}/lib
2 wget https://www.bioinformatics.babraham.ac.uk/projects/hicup/hicup_v0.6.1.tar.gz
3 tar -xzf hicup_v0.6.1.tar.gz
4 cd hicup_v0.6.1
5 chmod a+x *
```

2.3.4 SAMtools

```
1 cd ${HOME}/lib
2 git clone https://github.com/samtools/htslib.git
3 cd htslib
4 autoheader
5 autoconf
6 ./configure --prefix=${HOME}/lib/htslib
7 make
8 make install
9
```

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```

10 cd ${HOME}/lib
11 git clone https://github.com/samtools/samtools.git
12 cd samtools
13 autoheader
14 autoconf -Wno-syntax
15 ./configure --prefix=${HOME}/lib/samtools
16 make
17 make install

```

2.3.5 Install CHiCAGO

```

1 sudo apt-key adv --keyserver keyserver.ubuntu.com --recv-keys_
  ↪E298A3A825C0D65DFD57CBB651716619E084DAB9
2 sudo add-apt-repository 'deb [arch=amd64,i386] https://cran.rstudio.com/bin/linux/
  ↪ubuntu xenial/'
3 sudo apt-get update -qq
4 sudo apt-get install r-base-core
5 sudo apt-get install python-rpy2
6
7
8 cd ${HOME}/lib
9 sudo apt-get install libtbb-dev
10 sudo apt-get install libssl-dev
11 cd ${HOME}/C-HiC/
12 echo "R_LIB=${HOME}/R" > ${HOME}/.Renviro
13 echo "options(repos = c(CRAN = 'http://mirrors.ebi.ac.uk/CRAN/'))" > ${HOME}/.Rprofile
14 echo ".libPaths('~/.R') " >> ${HOME}/.Rprofile
15 echo 'message("Using library:", .libPaths()[1])' >> ${HOME}/.Rprofile
16 sudo Rscript CHiC/tool/scripts/install_packages.R
17
18 cd ${HOME}/C-HiC/CHiC/tool/scripts/
19 wget https://bitbucket.org/chicagoTeam/chicago/raw/
  ↪e288015f75d36c5367d1595e0ac8099f2ce82aa1/chicagoTools/runChicago.R
20 wget https://bitbucket.org/chicagoTeam/chicago/raw/
  ↪e288015f75d36c5367d1595e0ac8099f2ce82aa1/chicagoTools/bam2chicago.sh
21 wget https://bitbucket.org/chicagoTeam/chicago/raw/
  ↪e288015f75d36c5367d1595e0ac8099f2ce82aa1/chicagoTools/makeDesignFiles.py
22 chmod +x bam2chicago.sh

```

2.4 Setup the symlinks

```

1 cd ${HOME}/bin
2
3
4
5 ln -s ${HOME}/lib/hicup_v0.6.1/* ${HOME}/bin/
6
7 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2 bowtie2
8 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-align-s bowtie2-align-s
9 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-align-l bowtie2-align-l
10 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-build bowtie2-build
11 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-build-s bowtie2-build-s

```

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```
12 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-build-1 bowtie2-build-1
13 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-inspect bowtie2-inspect
14 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-inspect-s bowtie2-inspect-s
15 ln -s ${HOME}/lib/bowtie2-2.3.4-linux-x86_64/bowtie2-inspect-l bowtie2-inspect-l
16
17 ln -s ${HOME}/lib/htslib/bin/bgzip bgzip
18 ln -s ${HOME}/lib/htslib/bin/htsfile htsfile
19 ln -s ${HOME}/lib/htslib/bin/tabix tabix
20
21
22 ln -s ${HOME}/lib/samtools/bin/ace2sam ace2sam
23 ln -s ${HOME}/lib/samtools/bin/blast2sam.pl blast2sam.pl
24 ln -s ${HOME}/lib/samtools/bin/bowtie2sam.pl bowtie2sam.pl
25 ln -s ${HOME}/lib/samtools/bin/export2sam.pl export2sam.pl
26 ln -s ${HOME}/lib/samtools/bin/interpolate_sam.pl interpolate_sam.pl
27 ln -s ${HOME}/lib/samtools/bin/maq2sam-long maq2sam-long
28 ln -s ${HOME}/lib/samtools/bin/maq2sam-short maq2sam-short
29 ln -s ${HOME}/lib/samtools/bin/md5fa md5fa
30 ln -s ${HOME}/lib/samtools/bin/md5sum-lite md5sum-lite
31 ln -s ${HOME}/lib/samtools/bin/novo2sam.pl novo2sam.pl
32 ln -s ${HOME}/lib/samtools/bin/plot-bamstats plot-bamstats
33 ln -s ${HOME}/lib/samtools/bin/psl2sam.pl psl2sam.pl
34 ln -s ${HOME}/lib/samtools/bin/sam2vcf.pl sam2vcf.pl
35 ln -s ${HOME}/lib/samtools/bin/samtools samtools
36 ln -s ${HOME}/lib/samtools/bin/samtools.pl samtools.pl
37 ln -s ${HOME}/lib/samtools/bin/seq_cache_populate.pl seq_cache_populate.pl
38 ln -s ${HOME}/lib/samtools/bin/soap2sam.pl soap2sam.pl
39 ln -s ${HOME}/lib/samtools/bin/varfilter.py varfilter.py
40 ln -s ${HOME}/lib/samtools/bin/wgsim wgsim
41 ln -s ${HOME}/lib/samtools/bin/wgsim_eval.pl wgsim_eval.pl
42 ln -s ${HOME}/lib/samtools/bin/zoom2sam.pl zoom2sam.pl
```

2.5 Prepare the Python Environment

2.5.1 Install APIs and Pipelines

Checkout the code for the DM API and the C-HiC pipelines:

```
1 cd ${HOME}/code
2 pyenv activate C-HiC
3 pip install --upgrade setuptools pip
4 pip install git+https://github.com/Multiscale-Genomics/mg-dm-api.git
5 pip install git+https://github.com/Multiscale-Genomics/mg-tool-api.git
6 pip install git+https://github.com/Multiscale-Genomics/mg-process-fastq.git
7
8 git clone https://github.com/pabloacera/C-HiC.git
9 cd C-HiC
10 pip install -e .
11 pip install -r requirements.txt
12 pip install dill
```

3.1 Map and parse CHi-C reads

This pipeline will take as input two fastq files, RE sites, the genome indexed with GEM and the same genome in FASTA file. This pipeline uses TADbit to map, filter and produce a bed file that will be used later on to produce bam file compatible with CHiCAGO algorithm. More information about filtering and mapping <https://3dgenomes.github.io/TADbit/>

3.1.1 Running from the command line

Parameters

config [str] Configuration JSON file

in_metadata [str] Location of input JSON metadata for files

out_metadata [str] Location of output JSON metadata for files

Returns

Wd [folders and files] path to the working directory where the output files are

Example

REQUIREMENT - Needs two fastq files single end, FASTA genome and bowtie2 indexed genome.

When running the pipeline on a local machine without COMPSs:

```
1 python process_hicup.py \  
2   --config tests/json/config_hicup.json \  
3   --in_metadata tests/json/input_hicup.json \  
4
```

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```
4 --out_metadata tests/json/output_hicup.json \  
5 --local
```

When using a local version of the [COMPS virtual machine](<https://www.bsc.es/research-and-development/software-and-apps/software-list/comp-superscalar/>):

```
1 runcompss \  
2 --lang=python \  
3 --library_path=${HOME}/bin \  
4 --pythonpath=/<pyenv_virtenv_dir>/lib/python2.7/site-packages/ \  
5 --log_level=debug \  
6 process_fastq2bed.py \  
7 --config tests/json/config_hicup.json \  
8 --in_metadata tests/json/input_hicup.json \  
9 --out_metadata tests/json/output_hicup.json
```

3.1.2 Methods

class `process_hicup.process_hicup` (*configuration=None*)

This class run hicup tool which run hicup, doing the alignment and filtering of the reads and convert them into a BAM file.

run (*input_files, metadata, output_files*)

This is the main function that runs

Parameters

- **input_files** (*dict*) – fastq1 fastq2
- **metadata** (*dict*) –
- **output_files** (*dict*) –
- **out_dir**: **str** directory to write the output

Returns

- **results** (*bool*)
- **output_metadata** (*dict*)

3.2 Create CHiCAGO input RMAP

3.3 Create CHiCAGO input BAITMAP

3.4 Create CHiCAGO input Design files

This script use as input .rmap and .baitmap files and generate the Design files. NPerBin file (.npb): <baitID> <Total no. valid restriction fragments in distance bin 1> ... <Total no. valid restriction fragments in distance bin N>, where the bins map within the “proximal” distance range from each bait (0; maxLBrownEst] and bin size is defined by the binsize parameter. NBaitsPerBin file (.nbpb): <otherEndID> <Total no. valid baits in distance bin 1> ... <Total no. valid baits in distance bin N>, where the bins map within the “proximal” distance range from each other end (0; maxLBrownEst] and bin size is defined by the binsize parameter. Proximal Other End (ProxOE) file (.poe): <baitID> <otherEndID> <absolute distance> for all combinations of baits and other ends that map within the “proximal” distance range from

each other (0; maxLBrownEst]. Data in each file is preceded by a comment line listing the input parameters used to generate them.

3.4.1 Running from the command line

Parameters

config [str] Configuration JSON file

in_metadata [str] Location of input JSON metadata for files

out_metadata [str] Location of output JSON metadata for files

Returns

“nbpb” : .nbpb file “npb” : .npb file “poe” : .poe file

Example

REQUIREMENT - Needs RMAP and BAITMAP files

When running the pipeline on a local machine without COMPSs:

```
1 python process_design.py \
2   --config tests/json/config_design.json \
3   --in_metadata tests/json/input_design.json \
4   --out_metadata tests/json/output_design.json \
5   --local
```

When using a local version of the [COMPS virtual machine](<https://www.bsc.es/research-and-development/software-and-apps/software-list/comp-superscalar/>):

```
1 runcomps \
2   --lang=python \
3   --library_path=${HOME}/bin \
4   --pythonpath=/<pyenv_virtenv_dir>/lib/python2.7/site-packages/ \
5   --log_level=debug \
6   process_design.py \
7   --config tests/json/config_design.json \
8   --in_metadata tests/json/input_design.json \
9   --out_metadata tests/json/output_design.json
```

3.4.2 Methods

class process_design.**process_design** (*configuration=None*)

This class generates the Design files and chinput files, input for CHiCAGO. Starting from rmap and baitmap and capture HiC BAM files.

run (*input_files, metadata, output_files*)

Main function to run the tools, MakeDesignFiles_Tool.py and bam2chicago_Tool.py

Parameters

- **input_files** (*dict*) – designDir: path to the folder with .rmap and .baitmap files
rmapFile: path to the .rmap file baitmapFile: path to the .baitmap file bamFile: path to the capture HiC bamfiles
- **metadata** (*dict*) – input metadata
- **output_files** (*dict*) – outPrefixDesign : Path and name of the output prefix, recommend to be the same as rmap and baitmap files. sample_name: Path and name of the .chinput file

Returns

- *bool*
- *output_metadata*

3.5 Convert BAM file into chicago input files .chinput

3.6 Data normalization and peak calling

This pipeline runs the normalization of the data and call the real chomatine interactions

3.6.1 Running from the command line

Parameters

config [str] Configuration JSON file

in_metadata [str] Location of input JSON metadata for files

out_metadata [str] Location of output JSON metadata for files

Returns

output_dir: directory with all output folders and files

Example

REQUIREMENT - Needs a reference genome

- **Needs file with the capture sequences with FASTA format**
 - settings file
 - **design dir:** .rmap .baitmap .npb .nbpb .poe

When running the pipeline on a local machine without COMPSs:

```
1 python process_run_chicago.py \  
2     --config tests/json/config_chicago.json \  
3     --in_metadata tests/json/input_chicago.json \  
4     --out_metadata tests/json/output_chicago.json \  
5     --local
```

When using a local version of the [COMPS virtual machine]([https://www.bsc.es/research-and-development/ software-and-apps/software-list/comp-superscalar/](https://www.bsc.es/research-and-development/software-and-apps/software-list/comp-superscalar/)):

```

1 runcompss \
2   --lang=python \
3   --library_path=${HOME}/bin \
4   --pythonpath=/<pyenv_virtenv_dir>/lib/python2.7/site-packages/ \
5   --log_level=debug \
6   process_runChicago.py \
7     --config tests/json/config_chicago.json \
8     --in_metadata tests/json/input_chicago.json \
9     --out_metadata tests/json/output_chicago.json

```

3.6.2 Methods

class `process_run_chicago.process_run_chicago` (*configuration=None*)

Function for processing capture Hi-C fastq files. Files are aligned, filtered and analysed for Cpature Hi-C peaks

run (*input_files, metadata, output_files*)

This main function that run the chicago pipeline with runChicago.R wrapper

Parameters

- **input_files** (*dict*) – location with the .chinput files. `chinput_file`: str in case there is one input file `chinput_file`: comma separated list in case there is more than one input file.
- **metadata** (*dict*) – Input metadata, str
- **output** (*dict*) – output file locations

Returns

- **output_files** (*dict*) – Folder location with the output files
- **output_metadata** (*dict*) – Output metadata for the associated files in `output_files`

3.7 Run the entire CHi-C pipeline

Tools for processing fastq C-HiC files

4.1 Map and parser reads

4.1.1 hicup_tool

class CHiC.tool.hicup_tool.**hicup** (*configuration=None*)

Tool to run hicup, from fastq to bam files

digest_genome (*genome_name, re_enzyme, genome_loc, re_enzyme2*)

This function takes a genome and digest it using a restriction enzyme specified

Parameters

- **genome_name** (*str*) – name of the output genome
- **re_enzyme** (*str*) – name of the enzyme used to cut the genome format example A[^]GATCT, BglII .
- **genome_loc** (*str*) – location of the genome in FASTA format
- **re_enzyme2** (*str*) – Restriction site 2 refers to the second, optional (other DNA shearing techniques such as sonication may be used) enzymatic digestion. This restriction site does NOT form a Hi-C ligation junction. This is the restriction enzyme that is used when the Hi-C sonication protocol is not followed. Typically the sonication protocol is followed.

static get_hicup_params (*params*)

Function to handle to extraction of commandline parameters and formatting them for use with hicup

Parameters **params** (*dict*) –

--bowtie	Specify the path to Bowtie
--bowtie2	Specify the path to Bowtie 2
--config	Specify the configuration file
--digest	Specify the digest file listing restriction fragment co-ordinates

--example	Produce an example configuration file
--format	Specify FASTQ format Options: Sanger, Solexa_Illumina_1.0, Illumina_1.3, Illumina_1.5
--help	Print help message and exit
--index	Path to the relevant reference genome Bowtie/Bowtie2 indices
--keep	Keep intermediate pipeline files
--longest	Maximum allowable insert size (bps)
--nofill	Hi-C protocol did NOT include a fill-in of sticky ends prior to ligation step and therefore FASTQ reads shall be truncated at the Hi-C restriction enzyme cut site (if present) sequence is encountered
--outdir	Directory to write output files
--quiet	Suppress progress reports (except warnings)
--shortest	Minimum allowable insert size (bps)
--temp	Write intermediate files (i.e. all except summaryfiles and files generated by HiCUP Deduplicator) to a specified directory
--threads	Specify the number of threads, allowing simultaneous processing of multiple files
--version	Print the program version and exit
--zip	Compress output

Returns

Return type list

hicup_alig_filt (*params, genome_digest, genome_index, genome_loc, fastq1, fastq2, outdir_tar*)

This function aligns the HiC read into a reference genome and filter them

Parameters

- **bowtie2_loc** –
- **genome_index** (*str*) – location of genome indexed with bowtie2
- **digest_genome** (*str*) – location of genome digested
- **fastq1** (*str*) – location of fastq1 file
- **fastq2** (*str*) – location of fastq2

Returns

Return type Bool

hicup_alig_filt_runner (***kwargs*)

This function runs the hicup_alig_filt

Parameters

- **bowtie2_loc** –
- **genome_index** (*str*) – location of genome indexed with bowtie2

- **digest_genome** (*str*) – location of genome digested
- **fastq1** (*str*) – location of fastq2 file
- **fastq2** (*str*) – location of fastq2

Returns**Return type** Bool**run** (*input_files*, *metadata*, *output_files*)

Function that runs and pass the parameters for all the functions

Parameters

- **input_files** (*dict*) –
- **metadata** (*dict*) –
- **output_files** (*dict*) –

untar_index (*genome_file_name*, *genome_idx*, *bt2_1_file*, *bt2_2_file*, *bt2_3_file*, *bt2_4_file*, *bt2_rev1_file*, *bt2_rev2_file*)

Extracts the Bowtie2 index files from the genome index tar file. :param genome_file_name: Location string of the genome fasta file :type genome_file_name: str :param genome_idx: Location of the Bowtie2 index file :type genome_idx: str :param bt2_1_file: Location of the <genome>.1.bt2 index file :type bt2_1_file: str :param bt2_2_file: Location of the <genome>.2.bt2 index file :type bt2_2_file: str :param bt2_3_file: Location of the <genome>.3.bt2 index file :type bt2_3_file: str :param bt2_4_file: Location of the <genome>.4.bt2 index file :type bt2_4_file: str :param bt2_rev1_file: Location of the <genome>.rev.1.bt2 index file :type bt2_rev1_file: str :param bt2_rev2_file: Location of the <genome>.rev.2.bt2 index file :type bt2_rev2_file: str

Returns Boolean indicating if the task was successful**Return type** bool

4.2 Create CHiCAGO input files

4.2.1 makeRmap

4.2.2 makeBaitmap

4.2.3 makeDesignFiles

class CHiC.tool.makeDesignFiles.**makeDesignFilesTool** (*configuration=None*)

Tool for making the design files as part of the input for Chicago capture Hi-C

static **get_design_params** (*params*)

This function handle chicago parameters, selecting the given ones and passing to the command line.

makeDesignFiles (***kwargs*)

make the design files and store it in the specify design folder. It is a wrapper of makeDesignFiles.py

Parameters

- **designDir** (*str*,) – Path to the folder with the output files(recommended the same folder as .map and .baitmap files).
- **parameters** (*dict*,) – list of parameter already selected by get_makeDesignFiles_params().

Returns

- *bool*
- **outFilePrefix** (*str*) – writes the output files in the defined location

run (*input_files*, *input_metadata*, *output_files*)

The main function to run makeDesignFiles.

Parameters

- **input_files** (*dict*) – designDir : path to the designDir containin .rmap and .baitmap files
 - **input_metadata** (*dict*) –
 - **output_files** (*dict*) –
- outFilePrefix** [path to the output folder and prefix name of files] example: “/folder1/folder2/prefixname”. Recommended to use the path to designDir and the same prefix as .rmap and .baitmap

Returns

- **output_files** (*dict*) – List of location for the output files.
- **output_metadata** (*dict*) – List of matching metadata dict objects.

4.3 Convert bam files into chicago input

4.3.1 bam2chicago

4.4 Normalize data and call C-HiC peaks

4.4.1 run_chicago

class CHiC.tool.run_chicago.ChicagoTool (*configuration=None*)

tool for running the CHiCAGO algorithm

chicago (***kwargs*)

Run and annotate the Capture-HiC peaks. Chicago will create 4 folders under the output_prefix data : output_index.Rds -> chicago data saved on Rds format output_index_params.txt -> parameters used to run Chicago output_index.export_format -> chicago output in the chosen format diag_plots : 3 plots to assest the quality of the output (see CHicago Capture-HiC documentation for details) enrichment_data: files for the feature enrichment output (in case is used) examples: output_index_proxExamples.pdf: random chosen peaks showing interactions regions see <http://regulatorygenomicsgroup.org/chicago> for more information

Parameters

- **input_files** (*str* ot *comma separated list if there is more than one replicate*) –
- **output_prefix** (*str*) –
- **output_dir** (*str* (*whole path for the output*)) –
- **params** (*dict*) –

Returns writes the output files in the defined location

Return type bool

static `get_chicago_params` (*params*)

Function to handle to extraction of commandline parameters and formatting them for use in the aligner for BWA ALN

Parameters `params` (*dict*) –

Returns

Return type list

run (*input_files*, *input_metadata*, *output_files*)

The main function to run chicago for peak calling. The input files are .chinput and are transformed from BAM files using bam2chicago.sh input files could be just one file or a comma separated files from more than one biological replicate. Technical replicates should be pooled to one .chinput

Parameters

- **input_files** (*dict*) – list of .chinput files, or str with a single .chinput file
- **input_metadata** (*dict*) –
- **output_files** (*dict with the output path*) –

Returns

- **output_files** (*Dict*) – List of locations for the output files,
- **output_metadata** (*Dict*) – List of matching metadata dict objects

static `untar_chinput` (*chinput_tar*)

This function take as input the tar chinput

Parameters `chinput_tar` (*str*) – path to the tar file, the tar files should have the same prefix name as the tar file

Returns

Return type list of untar files

5.1 25-09-2018 handling_chr_header branch merge with master

This rmap_tool.py from this branch take the chromosome format from the used the reference genome and output a file with two columns, dictionary like with number of the chromosome and the name of the chromosome from the reference genome. example 1 chr1 2 chr2 3 chr3 ect. . .

This file is passed to the makeBaitmap.py script and generate the .batimap file with the corresponding chromosome name. This is necessary as the rtrees used in makeBaitmap.py needs an integer instead of “chr” or any other format.

5.2 15-10-2018 mm_mods_for_makebaitmaps branch merge with master

This branch contains some modifications from Mark to solve issues with pyCOMPSs regarding makeBaitmap.py tool

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